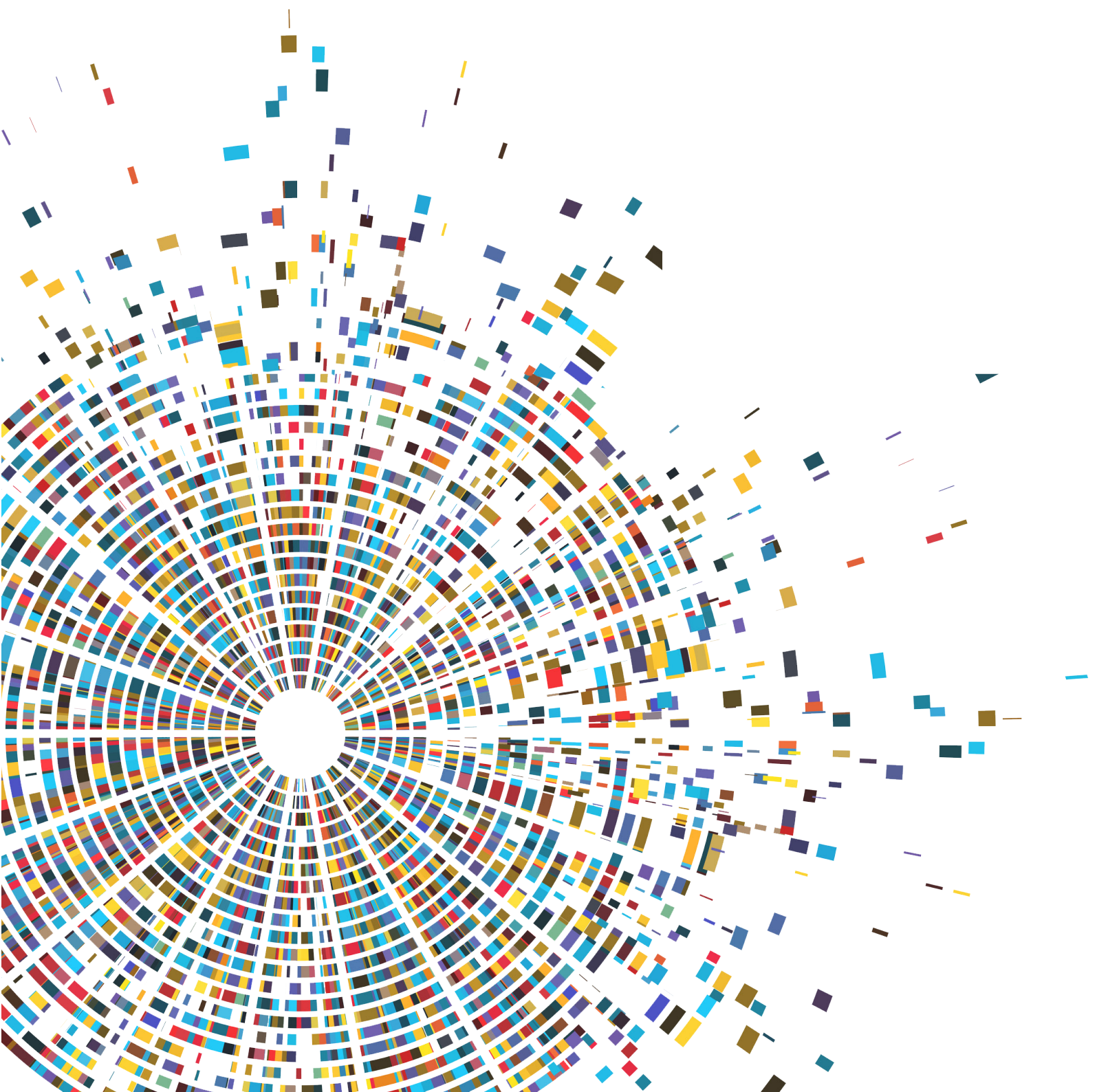




Genomic Unity[®] Test

A Whole Genome Clinical Validation Study



Overview

Variantix applies whole genome sequencing (WGS) in tandem with its proprietary Genomic Intelligence® analysis pipeline and software for the identification of clinically relevant variants. Nuclear WGS identifies single nucleotide variants (SNVs), small insertions and deletions (indels), structural variants (SVs) and short tandem repeat expansions (STRs), while mitochondrial genome sequencing identifies SNVs and heteroplasmy. This white paper describes the validation performed, providing the sensitivity and specificity for each variant type.

Background

Historically the detection of SNVs and indels have been performed using multiple sequencing platforms such as Sanger sequencing and Next Generation Sequencing (NGS), including whole exome and whole genome sequencing (WES/WGS) methodologies. However, in the broad scope of all genomic testing, this represents only a subset of methodologies used to identify pathogenic variants in the human genome. Other methodologies include: Southern blots to detect large STRs and large deletions, PCR and capillary electrophoresis to detect shorter STRs, qPCR and MLPA to detect deletions and duplications, and arrayCGH/microarray, FISH and karyotypes to detect gross chromosomal deletions.

A major concern in using multiple methods to detect different types of genetic variants is that one might miss a connection in a patient with a combination of changes that were identified by different method platforms at different times and often at different laboratories. For example, detection of a heterozygous deletion in a recessive gene in one lab and a heterozygous pathogenic SNV in another lab might be reported as “carrier” by each lab. To assimilate these independent lines of evidence into a positive result requires a comprehensive understanding of molecular biology and the complexities of genetic testing. This connection might be missed, even by a technically savvy ordering clinician.

Moreover, since each test has its own turn-around-time and is often ordered sequentially as a reflex off a prior result for financial reasons, disjointed tests offered at different labs place a burden on the family for multiple sample collections as well as extending the time to diagnosis, which can delay clinical management implementation. Therefore, consolidating previous multi-platform testing into one unified report is highly desirable in the clinical genomics diagnostic space to improve diagnostic yields, shorten patient turnaround times and provide the earliest possible window for clinical management and intervention.

With its PCR free DNA preparation and consistent read depth, WGS provides the opportunity for such a consolidated test. At Variantix, we have developed the Genomic Unity® test to provide comprehensive analysis of the variant types for which validation is described in this document.

Methodology

30X whole genome sequencing was performed on the Illumina platform using the Illumina TruSeq PCR Free DNA Preparation Kit. Data analysis was performed with version 2.7.0.0 of Variantix’s Genomic Intelligence® analysis pipeline and software.

Whenever possible, validation was performed using NIST Genome in a Bottle (GIAB) reference datasets. On average, about 1.4% of the genome does not have coverage sufficient for reliable variant calling (<8X), however only about 0.4% of clinically relevant variants (based on HGMD Professional and ClinVar annotations) are not adequately covered. See the Appendix for coverage and sequence depth metrics for GIAB samples. When acceptable “gold standard” true positive data sets did not exist, as in the case of structural variants and short tandem repeats, sourced controls were used as described.

Single nucleotide variants (SNVs)

Analysis of SNVs is performed using GATK best practice guidelines. Validation was performed on three GIAB samples: NA24149, NA24143 and NA12878. The reported validation statistics are based on the mean of true/false positive/negative values calculated for the three samples.

True positive (TP)	2,723,422
False positive (FP)	4,709
True negative (TN)	2,188,155,955
False negative (FN)	3,146
Sensitivity	0.99885
Specificity	1.00000
Positive predictive value	0.99827
Accuracy	1.00000

The following tables provide validation statistics for SNVs divided into heterozygous and homozygous (reference and alternate alleles combined) calls.

Heterozygous SNVs

True positive (TP)	1,656,415
False positive (FP)	4,648
True negative (TN)	2,189,182,961
False negative (FN)	2,573
Sensitivity	0.99846
Specificity	1.00000
Positive predictive value	0.99720
Accuracy	1.00000

Homozygous SNVs

True positive (TP)	1,067,006
False positive (FP)	61
True negative (TN)	2,189,772,370
False negative (FN)	573
Sensitivity	0.99946
Specificity	1.00000
Positive predictive value	0.99994
Accuracy	1.00000

Sensitivity (>99.8%), specificity (100%), positive predictive value (>99.7%) and accuracy (100%) of SNVs called by Genomic Unity® test are at or above those typically reported for whole exome sequencing by multiple clinical laboratories.

Small insertions and deletions (indels)

Analysis of indels is performed using GATK best practice guidelines. Validation was performed on three GIAB samples: NA24149, NA24143 and NA12878. The reported validation statistics are grouped by indel size and based on the mean of true/false positive values calculated for the three samples. Indel sizes of 1-4 bp, 5-15 bp and ≥16 bp are considered. However, there is not a hard size limit as the largest indel included was 172 bp.

1-4 bp

True positive (TP)	389,023
False positive (FP)	4,917
Sensitivity	0.98552

5-15 bp

True positive (TP)	31,393
False positive (FP)	546
Sensitivity	0.98384

≥16 bp

True positive (TP)	6,187
False positive (FP)	128
Sensitivity	0.97176

The following tables provide validation statistics for indels divided into heterozygous and homozygous (reference and alternate alleles combined) calls.

Heterozygous indels

1-4 bp

True positive (TP)	242,396
False positive (FP)	2,425
Sensitivity	0.98685

5-15 bp

True positive (TP)	20,677
False positive (FP)	245
Sensitivity	0.98074

≥16 bp

True positive (TP)	4,200
False positive (FP)	78
Sensitivity	0.96772

Homozygous indels

1-4 bp

True positive (TP)	146,627
False positive (FP)	2,492
Sensitivity	0.98332

5-15 bp

True positive (TP)	10,716
False positive (FP)	301
Sensitivity	0.98985

≥16 bp

True positive (TP)	1,987
False positive (FP)	50
Sensitivity	0.98042

Sensitivity of the Genomic Unity® test in calling of indels of any size is at or above 96%.

Mitochondrial heteroplasmy

30X whole genome sequencing produces approximately 2,000X coverage of the mitochondrial genome. Mitochondrial variants were detected in 15 samples obtained by mixing DNA of two known samples in different ratios and sequencing the mixes. Greater than 1,000X coverage was observed at every nucleotide in all tested samples.

The following table shows how the percentage of alternate reads detected changes as the composition of the sequencing mixes progress from containing 0% to 100% of the second known sample. For each case, the listed value is the percentage of alternate reads detected out of the total

number of reads, which reflects the heteroplasmy of the called variants. In some cases, both samples are homozygous reference, consistently producing a score of 0% (for example, chrM:10925T>G). In other cases, both samples are homozygous alternate, consistently producing a score of 100% (for example, chrM:750A>G). It is the cases where one or both samples are heterozygous, and in some cases at varying degrees of heteroplasmy, that provide the greatest insight into the detectable level of heteroplasmy (for example, chrM:6776T>C). Note that in cases where the variant is not called due to the fraction of alternate reads falling below the calling threshold, the number of alternate reads is defaulted to 0 in turn producing a value of 0% regardless of the actual number of supporting reads.

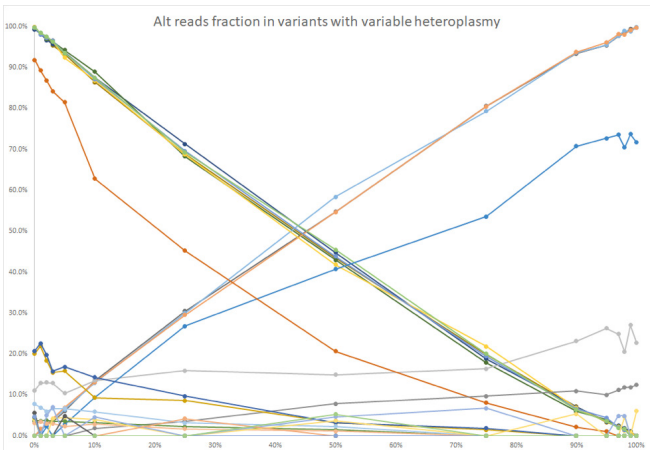
Variant name	0%	1%	2%	3%	5%	10%	25%	50%	75%	90%	95%	97%	98%	99%	100%
chrM:750A>G	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%
chrM:15326A>G	99.9%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	99.9%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%
chrM:16519T>C	100.0%	99.9%	100.0%	100.0%	99.9%	99.9%	100.0%	99.9%	100.0%	100.0%	99.9%	99.9%	100.0%	100.0%	100.0%
chrM:310T>TC	100.0%	99.9%	100.0%	99.9%	99.5%	99.5%	99.5%	100.0%	99.6%	99.7%	100.0%	99.8%	99.7%	100.0%	99.9%
chrM:1438A>G	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%
chrM:8860A>G	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	99.9%	100.0%	100.0%	100.0%	99.9%	100.0%
chrM:263A>G	99.8%	100.0%	100.0%	99.8%	100.0%	100.0%	100.0%	100.0%	99.9%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%
chrM:4769A>G	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	99.9%	100.0%	100.0%	100.0%
chrM:10535T>C	0.0%	1.5%	2.8%	3.4%	6.1%	13.5%	30.5%	54.8%	80.6%	93.4%	95.5%	97.7%	98.4%	99.5%	99.7%
chrM:6293T>A	99.5%	98.4%	97.3%	95.4%	93.3%	86.4%	68.9%	43.6%	19.8%	7.1%	3.3%	2.3%	1.4%	0.8%	0.0%
chrM:200A>G	99.3%	98.3%	96.7%	95.8%	93.7%	87.5%	71.3%	44.8%	18.8%	7.0%	3.4%	2.4%	1.9%	1.0%	0.0%
chrM:8602T>C	99.9%	98.3%	97.0%	96.4%	94.3%	89.0%	68.3%	43.0%	17.9%	6.0%	4.0%	2.3%	1.6%	0.9%	0.0%
chrM:15314G>A	0.0%	1.0%	2.9%	4.1%	6.4%	12.9%	30.0%	58.4%	79.3%	93.6%	95.5%	97.8%	99.0%	98.8%	99.9%
chrM:6776T>C	0.0%	1.7%	2.7%	4.1%	6.9%	13.1%	29.5%	54.7%	80.5%	93.8%	96.1%	98.2%	98.0%	99.2%	99.8%
chrM:302A>AC	11.1%	12.9%	13.1%	12.9%	10.4%	13.5%	15.9%	14.9%	16.4%	23.2%	26.3%	24.9%	20.5%	27.0%	22.7%
chrM:14212T>C	99.9%	98.2%	97.4%	96.6%	92.4%	86.9%	68.8%	41.8%	21.8%	6.8%	3.6%	2.3%	1.3%	0.8%	0.0%
chrM:8937T>C	99.8%	98.1%	97.3%	96.6%	93.6%	87.2%	69.6%	43.9%	19.4%	6.6%	4.4%	2.1%	1.4%	0.0%	0.0%
chrM:3010G>A	99.8%	98.4%	97.6%	96.4%	93.6%	87.5%	69.3%	45.5%	20.1%	6.4%	3.6%	2.0%	1.7%	0.0%	0.0%

Variant name (continued)	0%	1%	2%	3%	5%	10%	25%	50%	75%	90%	95%	97%	98%	99%	100%
chrM:302A>ACC	0.0%	0.0%	2.3%	0.0%	2.8%	9.4%	26.8%	40.7%	53.6%	70.8%	72.8%	73.6%	70.5%	73.8%	71.7%
chrM:16189T>C	91.8%	89.3%	86.8%	84.2%	81.5%	62.8%	45.3%	20.7%	8.1%	2.2%	1.1%	0.0%	0.0%	0.0%	0.0%
chrM:9053G>A	0.0%	0.0%	0.0%	0.0%	0.0%	1.8%	3.5%	7.8%	9.7%	10.9%	10.0%	11.1%	11.8%	11.9%	12.4%
chrM:16183A>ACC	20.1%	21.9%	18.3%	15.5%	15.8%	9.3%	8.6%	3.1%	1.5%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:16183A>AC	20.7%	22.6%	19.8%	15.8%	16.8%	14.3%	9.7%	3.2%	1.9%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10785T>C	3.6%	3.6%	3.8%	3.5%	3.5%	3.2%	2.3%	1.4%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:16188CT>C	7.8%	6.9%	6.0%	6.7%	6.6%	5.9%	3.2%	2.3%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:7162G>A	3.1%	3.4%	3.0%	3.0%	2.3%	2.9%	1.7%	1.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:1552G>A	1.7%	1.8%	1.8%	1.7%	1.8%	1.6%	1.6%	0.8%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:13762T>G	4.0%	0.0%	0.0%	4.2%	4.5%	3.9%	0.0%	3.7%	0.0%	5.3%	0.0%	0.0%	4.6%	0.0%	6.1%
chrM:539T>A	0.0%	0.0%	4.9%	7.1%	0.0%	4.6%	0.0%	4.6%	6.8%	0.0%	0.0%	4.9%	4.9%	0.0%	0.0%
chrM:814A>G	0.8%	0.7%	0.0%	0.0%	0.0%	0.0%	0.7%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10933C> CGGGGGGGGG	0.0%	0.0%	0.0%	0.0%	0.0%	2.4%	0.0%	0.0%	0.0%	0.7%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10569G>A	0.0%	0.7%	0.0%	0.0%	0.7%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10935A>G	5.7%	0.0%	0.0%	0.0%	4.8%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:13768T>G	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	3.1%	0.0%	0.0%
chrM:301A>C	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	3.9%	0.0%	0.0%
chrM:10936C>G	4.5%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:3776G>A	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.8%	0.0%
chrM:3577A>C	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	4.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10926T>G	0.0%	0.0%	0.0%	0.0%	0.0%	2.1%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:302A>ACCC	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	20.5%	0.0%	0.0%
chrM:10941T>G	4.6%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:484A>C	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	5.2%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%
chrM:10925T>G	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.7%	0.0%	0.0%	0.0%	0.0%	0.0%

The data from the table on the previous page is displayed graphically in the figure to the right.

The following table shows the same data, expressed as the number of reference and alternate reads (displayed as reference;alternate) measured in the sample in order to provide full sight into the coverage at each variant site.

Note that in cases where the variant is not called due to the fraction of alternate reads falling below the calling threshold, the value is defaulted to 0;0 regardless of the actual number of supporting reads.



Variant name	0%	1%	2%	3%	5%	10%	25%	50%	75%	90%	95%	97%	98%	99%	100%
chrM:750A>G	0;1706	0;1718	0;1845	1;2539	0;2569	0;2018	0;1995	0;2217	0;1872	0;1689	0;2285	0;1620	0;1860	0;1714	0;2753
chrM:15326A>G	1;1525	1;2369	0;2608	0;1549	0;2245	0;1557	0;1845	1;1461	0;1480	0;2064	0;1587	0;1467	0;2654	0;1497	0;1469
chrM:16519T>C	0;1759	1;1582	0;1902	0;1537	1;1494	1;1801	0;1451	1;1717	0;1748	0;2262	1;1817	1;1805	0;3758	0;1783	0;1419
chrM:310T>TC	0;763	1;787	0;861	1;777	4;779	5;997	4;826	0;861	3;724	3;871	0;848	2;895	3;1069	0;912	1;777
chrM:1438A>G	1;2045	0;2879	0;2649	0;2709	0;2934	0;2064	0;2591	0;2530	0;3172	1;3359	0;2175	0;3176	0;3094	0;3091	0;2486
chrM:8860A>G	0;1837	0;1743	0;1648	0;1701	0;1753	0;1498	0;1751	0;1612	0;1707	1;1765	0;1713	0;1539	0;1638	1;1966	0;2204
chrM:263A>G	2;1297	0;1321	0;1435	3;1748	0;942	0;1143	0;990	0;952	1;1248	0;1096	0;1027	0;1056	0;1415	0;1484	0;895
chrM:4769A>G	0;2340	0;1777	0;2250	0;2134	0;2753	0;2252	0;2098	0;3009	0;2204	0;1587	0;1887	1;1966	0;2015	0;2922	0;1961
chrM:10535T>C	0;0	2482;37	2406;70	2700;94	2340;151	2688;419	1440;631	1461;1768	384;1596	179;2543	84;1765	75;3207	46;2794	10;1904	6;2376
chrM:6293T>A	10;2025	43;2663	70;2484	93;1948	151;2094	369;2345	908;2013	1949;1504	1733;428	3424;262	3458;118	3871;90	3838;56	3604;30	0;0
chrM:200A>G	22;3226	57;3359	111;3255	131;2959	222;3322	311;2176	1082;2691	2054;1665	2719;630	3456;261	3779;135	3772;94	3851;76	3335;35	0;0
chrM:8602T>C	1;1391	32;1832	72;2340	90;2404	116;1903	308;2483	601;1294	1540;1163	1601;348	1589;101	1474;61	2621;62	2948;49	2489;23	0;0
chrM:15314G>A	0;0	2504;26	2409;71	1513;64	2239;154	1345;200	1429;612	628;882	312;1196	131;1924	82;1736	34;1482	24;2397	22;1750	2;1510
chrM:6776T>C	0;0	2358;40	2382;66	2479;105	2356;174	2001;302	1868;782	1158;1398	512;2118	132;1993	110;2723	53;2875	55;2765	22;2740	4;1821
chrM:302A>AC	649;81	642;95	679;102	649;96	628;73	736;115	491;93	399;70	281;55	212;64	188;67	193;64	0;0	181;67	177;52
chrM:14212T>C	2;1401	28;1531	38;1418	70;1972	145;1768	182;1208	552;1215	1274;915	1082;302	1583;116	1349;50	1387;32	1625;22	1501;12	0;0
chrM:8937T>C	5;2045	37;1898	42;1520	64;1823	126;1838	215;1467	564;1292	1080;844	1358;327	1967;139	2054;95	1950;41	1788;26	0;0	0;0
chrM:3010G>A	6;2910	49;3085	38;1532	114;3013	183;2665	412;2874	629;1417	1168;975	1906;479	2239;152	2604;98	2312;46	1489;26	0;0	0;0

Variant name (continued)	0%	1%	2%	3%	5%	10%	25%	50%	75%	90%	95%	97%	98%	99%	100%
chrM:302A>ACC	0;0	0;0	679;16	0;0	628;18	736;76	491;180	399;274	281;324	212;514	188;502	193;538	267;637	181;509	177;448
chrM:16189T>C	54;606	71;590	92;605	107;572	127;559	301;509	560;463	862;225	1285;113	1354;30	1402;15	0;0	0;0	0;0	0;0
chrM:9053G>A	0;0	0;0	0;0	0;0	0;0	2108;39	1553;57	1916;163	2029;217	1350;166	1549;173	1539;193	1893;254	1530;206	1488;211
chrM:16183A>ACC	428;108	417;117	463;104	480;88	494;93	634;65	817;77	991;32	1266;19	0;0	0;0	0;0	0;0	0;0	0;0
chrM:16183A>AC	428;112	417;122	463;114	480;90	494;100	634;106	817;88	991;33	1266;24	0;0	0;0	0;0	0;0	0;0	0;0
chrM:10785T>C	2078;78	2459;92	1479;58	2024;74	2439;89	2501;84	2605;60	3170;46	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:16188CT>C	616;52	624;46	662;42	641;46	661;47	772;48	998;33	1071;25	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:7162G>A	2705;87	2335;83	2061;63	2828;87	2370;56	2506;74	2184;38	2312;27	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:1552G>A	2132;36	3029;55	3084;56	2933;50	2943;53	3358;56	3195;53	3461;29	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:13762T>G	988;41	0;0	0;0	1083;48	1464;69	1023;42	0;0	927;36	0;0	1014;57	0;0	0;0	966;47	0;0	1134;74
chrM:539T>A	0;0	0;0	1083;56	1091;83	0;0	1082;52	0;0	1087;53	1750; 127	0;0	0;0	1034;53	1125;58	0;0	0;0
chrM:814A>G	1460;12	1756;13	0;0	0;0	0;0	0;0	1836;13	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:10933C> CGGGGGGGGG	0;0	0;0	0;0	0;0	0;0	1671;41	0;0	0;0	0;0	821;6	0;0	0;0	0;0	0;0	0;0
chrM:10569G>A	0;0	2633;18	0;0	0;0	2484;18	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:10935A>G	1110;67	0;0	0;0	0;0	1197;61	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:13768T>G	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	983;31	0;0	0;0
chrM:301A>C	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	1170;48	0;0	0;0
chrM:10936C>G	1105;52	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:3776G>A	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	1447;11	0;0
chrM:3577A>C	0;0	0;0	0;0	0;0	0;0	0;0	1724;75	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:10926T>G	0;0	0;0	0;0	0;0	0;0	1737;37	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:302A>ACCC	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	267;69	0;0	0;0
chrM:10941T>G	1014;49	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:484A>C	0;0	0;0	0;0	0;0	0;0	0;0	0;0	957;53	0;0	0;0	0;0	0;0	0;0	0;0	0;0
chrM:10925T>G	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	0;0	848;6	0;0	0;0	0;0	0;0	0;0

This assay demonstrates the ability of the Genomic Unity[®] test to correctly identify mitochondrial variants down to 5% heteroplasmy.

This study does not present validation for reproducible calling of large deletions of the mitochondrial genome, due to too small of a number of available controls. However, it should be noted that both CAP proficiency large mitochondrial deletion samples were accurately identified and reported (see structural variant table below).

Structural variants (SVs)

Analysis of SVs is performed using a proprietary combination of break point analysis of split and discordant reads and read depth analysis, which is described in detail in the publication by Neerman et al¹.

Unlike for SNVs and indels, no true positive “gold standard” variant set of acceptable quality is currently available for the secondary validation of SVs. The closest is the dataset published by Parikh et al² which uses the svclassify method to establish bench mark structural variant calls. Different sequencing and variant calling methods applied by different research groups produced sets of “true positive” variants which vary between each other by nearly an order of magnitude in terms of quantity and overlap of detected variants. Close examination of a representative group of “true positive” SVs identified by different approaches revealed large numbers of false positives and false negatives, therefore making use of this data less than ideal for analytical validation of this clinical test. The statistics below are based on the available true positive data set NA24385 and will be used until better quality true positive SV data becomes available.

CNV <1,000	True positive (TP)	1,586	False negative (FN)	472
	False positive (FP)	574	Sensitivity	0.77100
CNV 1,000-10,000	True positive (TP)	486	False negative (FN)	82
	False positive (FP)	311	Sensitivity	0.85600
CNV 10,000-100,000	True positive (TP)	27	False negative (FN)	11
	False positive (FP)	111	Sensitivity	0.71000
CNV >100,000	True positive (TP)	1	False negative (FN)	0
	False positive (FP)	0	Sensitivity	1.00000

To more accurately reflect the true sensitivity and specificity of the algorithms, we analyzed known structural variants which are summarized in the table below. The following data set comprises true positive (causative pathogenic SVs confirmed by orthogonal detection techniques) and true negative clinical samples (those of healthy individuals or affected individuals with another type of causative genetic variant). Most were obtained from public collections, while some originated from other sources. This data set is discussed in more detail in the publication by Neerman et al¹.

Type of variant (Size, bp)	Variantyx detected (Size, bp)
24 variants including 2 SVs (126bp, 343bp)	Yes (126), Yes (break point)
Trisomy Chr 21	Yes (whole chromosome)
Trisomy Chr 18	Yes (whole chromosome)
Trisomy Chr 13	Yes (whole chromosome)
SNV + gross deletion (10kbp)	Yes (9,789 bp)
Compound het point mutation with large deletion, Krabbe disease, GALC (31kbp)	Yes (31,666 bp)
Deletion chrX male (8kbp)	Yes (8,749 bp)
Het deletion GGA compound with splicing point mutation (8kbp)	Yes (8,267 bp)
arr 22q11.22q11.23(21,390,449-21,978,719)x1	Yes (679,500 bp)
arr 2q37.2q37.3(236,218,793-242,654,701)x3,1 1q25(131,542,057-134,434,130)x1	Yes (6,484,000 bp), Yes (2,925,000 bp)

arr[hg19] 5q33.1q35.3(152,281,639-180,686,444)x3,7q36.2q36.3(155,040,999-159,123,167)x1	Yes (4,117,500 bp), Yes (337,000 bp, and few additional smaller duplication within the same locus)
Deletion DMD exon 44	Yes (195,012 bp)
Gross deletion (6 mbp)	Yes (5,979,236 bp)
Gross deletion (340 kbp)	Yes (340,153 bp)
45 bp deletion	Yes (45bp)
Gross duplication (700 kbp)	Yes (712,000 bp)
Chromosome deletion	Yes (16,095,000 bp)
Duplicated chromosome copy number variation (CNV) reference panel	Yes (17,983,000 bp)
Translocated chromosome	Yes (3,393,500 bp deletion; 8,112,000 bp duplication)
Recombinant chromosome copy number variantion (CNV) reference panel 02	Yes (1,108,000 bp deletion; 13,109,000 bp duplication)
Mitochondrial deletion	Yes (8,959 bp)
Mitochondrial deletion	Yes (7,647 bp)

The above samples include 18 SVs >100kb, of which all were detected, supporting a sensitivity and specificity of detection of large structural variants of 100%.

The Genomic Unity® test detects pathogenic SVs of multiple types with over 96% clinical sensitivity, with examples of reported pathogenic variants ranging from a 45 bp deletion to a complex rearrangement involving millions of base pairs on three different chromosomes.

Short tandem repeat expansions (STRs)

Analysis of STRs is performed using the application of three paired-end read strategies: de novo assembly of spanning reads, alignment and counting of repeats in anchored reads and counting and statistical normalization of reads containing only repeat sequence.

No true positive pathogenic STRs are available within GIAB samples, thus these types of variants were validated using samples from patients with confirmation of disease and a confirmed genetic diagnosis made by an orthogonal technology at CLIA-certified laboratories. Further details are found in the table below.

Sample	Gene (region)	Repeat unit	Reference count	Variantyx count	Reporting threshold
1	AR	CAG	51	52	35
2	ATXN1	CAG	15/65	19/68	36
3	ATXN1	CAG	29/52	30/61	36
4	ATXN1	CAG	31/43	32/38	36
5	ATXN1	CAG	16/68	20/67	36
6	ATXN1	CAG	32/60	33/69	36
7	ATXN2	CAG	Intermediate (27-33)	23/27	27
8	ATXN2	CAG	Intermediate (27-33)	23/27	27
9	ATXN2	CAG	Intermediate (27-33)	22/27	27
10	ATXN2	CAG	Intermediate (27-33)	22/31	27
11	ATXN2	CAG	Intermediate (27-33)	Intermediate 23/27	27

Sample	Gene (region)	Repeat unit	Reference count	Variantyx count	Reporting threshold
12	ATXN2	CAG	Intermediate (27-33)	Intermediate 23/27	27
13	ATXN2	CAG	Intermediate (27-33)	Intermediate 23/27	27
14	ATXN2	CAG	Intermediate (27-33)	Intermediate 22/31	27
15	ATXN3	CAG	24/74	57/63	27
16	ATXN7	CAG	8/62	7/76	27
17	C9ORF72	GGGGCC	Positive	2/1062	25
18	C9ORF72	GGGGCC	Positive	2/903	25
19	C9ORF72	GGGGCC	Positive	2/336	25
20	C9ORF72	GGGGCC	Positive C9Orf72 5/84/>145	Positive C9Orf72 5/176	25
21	C9ORF72	GGGGCC	Positive C9Orf72 5/84/>145	Positive C9Orf72 5/176	25
22	C9ORF72	GGGGCC	Positive C9Orf72 8/>145	Positive C9Orf72 8/268	25
23	C9ORF72	GGGGCC	Positive C9Orf72 8/>145	Positive C9Orf72 8/268	25
24	C9ORF72	GGGGCC	Positive C9Orf72 10/>145	Positive C9Orf72 10/114	25
25	C9ORF72	GGGGCC	Positive C9orf72 10/>145	Positive C9Orf72 10/114	25
26	C9ORF72	GGGGCC	Positive C9Orf72 4/>145	Positive C9Orf72 4/141	25
27	C9ORF72	GGGGCC	Positive C9Orf72 4/>145	Positive C9Orf72 4/141	25
28	C9ORF72	GGGGCC	Intermediate C9Orf72 11/28	Intermediate C9Orf72 11/32	25
29	C9ORF72	GGGGCC	Intermediate C9Orf72 11/28	Intermediate C9Orf72 11/32	25
30	DMPK	CTG	Normal/over 1500	14/2340	36
31	DMPK	CTG	Normal/over 1500	5/2255	36
32	DMPK	CTG	1000	5/1022	36
33	DMPK	CTG	500-1500	12/684	36
34	DMPK	CTG	500	532	36
35	DMPK	CTG	Normal/66	12/71	36
36	DMPK	CTG	5/1700	5/1661	36
37	DMPK	CTG	500-1000	12/689	36
38	DMPK	CTG	Normal/500	13/482	36
39	DMPK	CTG	Affected	12/572	36
40	DMPK	CTG	700	21/517	36
41	DMPK	CTG	340	21/288	36
42	DMPK	CTG	Normal/150-160	5/416	36
43	DMPK	CTG	Affected	14/119	36
44	DMPK	CTG	50-80	14/119	36
45	DMPK	CTG	1500-2000	13/3620	36
46	DMPK	CTG	Normal/80-90	22/127	36
47	DMPK	CTG	Normal/130-140	5/405	36
48	FMR1	CGG	30	30	45
49	FMR1	CGG	477	87	45
50	FMR1	CGG	80-93	67	45
51	FMR1	CGG	23	23	45
52	FMR1	CGG	20/29	20/29	45
53	FMR1	CGG	Affected	101	45
54	FMR1	CGG	23	23	45
55	FMR1	CGG	Affected	112	45
56	FMR1	CGG	23	23	45
57	FMR1	CGG	23/30	23/30	45
58	FMR1	CGG	Affected	174	45

Sample	Gene (region)	Repeat unit	Reference count	Variantyx count	Reporting threshold
59	FMR1	CGG	Affected	174	45
60	FMR1	CGG	32/107	33/73	45
61	FMR1	CGG	Affected	69	45
62	FMR1	CGG	23/95-140	23/80	45
63	FMR1	CGG	24,99	24/66	45
64	FMR1	CGG	29,69	29/56	45
65	FMR1	CGG	31,>200	31/113	45
66	FMR1	CGG	34,>200	34/98	45
67	FMR1	CGG	69	68	45
68	FMR1	CGG	>200	81	45
69	FMR1	CGG	>200	91	45
70	FMR1	CGG	20/183-193	20/74	45
71	FMR1	CGG	30/80	59/70	45
72	FMR1	CGG	31/46	31/54	45
73	FMR1	CGG	29/93-110	29/66	45
74	FMR1	CGG	Affected	105	45
75	FMR1	CGG	30/75-89	30/71	45
76	FMR1	CGG	76	56	45
77	FMR1	CGG	Affected	97	45
78	FMR1	CGG	931-940	117	45
79	FMR1	CGG	29/41	29/29	45
80	FMR1	CGG	53	58	45
81	FMR1	CGG	23/30	23/30	45
82	FMR1	CGG	46	54	45
83	FMR1	CGG	645	115	45
84	FMR1	CGG	29/29	29/29	45
85	FMR1	CGG	100-104	71	45
86	FMR1	CGG	23/30	23/30	45
87	FMR1	CGG	23/95	24/60	45
88	FMR1	CGG	Affected	126	45
89	FMR1	CGG	29/31	29/31	45
90	FMR1	CGG	80-96	63	45
91	FMR1	CGG	501-550	100	45
92	FMR1	CGG	29/30	29/30	45
93	FMR1	CGG	100-118	80	45
94	FMR1	CGG	Affected	127	45
95	FMR1	CGG	23	23	45
96	FMR1	CGG	Affected	87	45
97	FMR1	CGG	41	35	45
98	FMR1	CGG	>200	89	45
99	FMR1	CGG	31/53	31/52	45
100	FMR1	CGG	30	30	45
101	FMR1	CGG	23/29	24/29	45
102	FMR1	CGG	29/45	29/40	45
103	FMR1	CGG	23/29	23/29	45
104	FMR1	CGG	30/73	30/57	45
105	FMR1	CGG	Carrier	32/107	45

Sample	Gene (region)	Repeat unit	Reference count	Variantyx count	Reporting threshold
106	FMR1	CGG	30/78	30/54	45
107	FMR1	CGG	117	70	45
108	FMR1	CGG	23/70	23/56	45
109	FMR1	CGG	29/over 200	29/71	45
110	FXN	GAA	670/830	22/169	60
111	FXN	GAA	670/830	6/193	60
112	FXN	GAA	Normal/830	8/90	60
113	FXN	GAA	Normal/670	19/96	60
114	FXN	GAA	200/500	116/186	60
115	FXN	GAA	760/830	113/178	60
116	FXN	GAA	Normal/700	8/84	60
117	FXN	GAA	Normal/830	9/112	60
118	FXN	GAA	800/800	102/155	60
119	FXN	GAA	280/830	8/183	60
120	FXN	GAA	500	8/113	60
121	FXN	GAA	Normal/760	9/111	60
122	FXN	GAA	600/700	100/150	60
123	FXN	GAA	330/380	97/146	60
124	FXN	GAA	420/541	99/147	60
125	FXN	GAA	Normal/420	8/94	60
126	FXN	GAA	580/580	112/174	60
127	FXN	GAA	Normal/830	79/109	60
128	FXN	GAA	650/1030	111/175	60
129	FXN	GAA	Normal/830	9/101	60
130	FXN	GAA	9/1285+50	9/85	60
131	FXN	GAA	670/1170	116/183	60
132	FXN	GAA	630/830	109/168	60
133	FXN	GAA	Normal/830	8/103	60
134	FXN	GAA	530/530	98/146	60
135	HTT	CAG	17/48	17/61	36
136	HTT	CAG	15/52	15/58	36
137	HTT	CAG	22/58	22/63	36
138	HTT	CAG	16/66	16/82	36
139	HTT	CAG	15/70	15/78	36
140	HTT	CAG	22/50	22/60	36
141	HTT	CAG	16/44	16/52	36
142	HTT	CAG	45/47	43/56	36
143	HTT	CAG	15/49	15/62	36
144	HTT	CAG	17/45	17/43	36
145	HTT	CAG	16/46	16/52	36
146	HTT	CAG	15/55	15/61	36
147	HTT	CAG	15/44	15/58	36

The Genomic Unity® test detects pathogenic STRs of multiple types with over 99% clinical sensitivity.

Repeat variance was calculated for each expansion type:

CAG / CTG repeats (reverse allele, includes DMPK)

The following genes in this data set have CAG repeat expansions: HTT, ATXN1, ATXN2, ATXN3, ATXN7, AR.

The variance between actual and detected CAG repeats from 1-40 in size is +/- 1.

The variance between actual and detected CAG repeats from >40 in size is +/- 20%.

GAA repeats

The following gene in this data set has a GAA repeat expansion: FXN.

FXN GAA repeats detected as 1-60 in size are reported as “normal”. The detected repeat size is reported with a variance of +/- 10%.

Due to the abundance of naturally occurring GAA repeats in the genome, repeats >60 in size are reported as “heterozygous” or “homozygous” based on the correlation between detected and actual repeat size determined during validation. Repeats detected as 61-140 in size are indicative of carrier status due to heterozygous expansion of one allele and are reported as “heterozygous”. Repeats detected as >140 in size are indicative of a homozygous or compound heterozygous expansion of both alleles and are reported as “homozygous”.

Orthogonal confirmation is required to determine exact allele sizes of >60.

GGGGCC repeats

The following gene in this data set has a GGGGCC repeat expansion: C9orf72.

The variance between actual and detected GGGGCC repeats from 1-30 in size is +/- 1.

The variance between actual and detected GGGGCC repeats >30 in size is +/- 20%. Note that the variance for repeats >30 in size is challenging to determine as large expansions called with orthogonal technologies do not provide exact repeat counts.

CGG repeats

The following gene in this data set has a CGG repeat expansion: FMR1.

FMR1 CGG repeats detected as 1-55 in size are reported as “normal”. The detected repeat size is reported with a variance of +/- 1.

FMR1 CGG repeats detected as >55 in size are reported as “pre-mutation or full mutation”. These expansions are not reported as an exact number of repeats due to the challenging nature of sequencing and aligning this region.

Orthogonal confirmation is recommended to determine whether the repeat is of pre-mutation (56-200) or full mutation (>200) size.

Conclusion

This validation document provides data supporting the ability to reliably identify clinically relevant SNVs, indels, mitochondrial variants, SVs, and STRs using whole genome sequencing and Variantyx’s proprietary Genomic Intelligence® software.

References

1. Neerman et al. A clinically validated whole genome pipeline for structural variant detection and analysis. BMC Genomics. 2019 Jul 16;20(Suppl 8):545.
2. Parikh H et al. svclassify: A method to establish benchmark structural variant calls. BMC Genomics. 2016 Jan 16;17:64.

Appendix

The following table provides coverage and sequence depth metrics for three GIAB samples across all loci.

	NA24149	NA24143	NA12878
Total coverage	118,834,360,234	104,452,111,795	103,137,319,942
Total bases	2,937,658,094	2,937,658,094	2,937,658,094
<20x coverage bases	141,669,917	116,038,970	128,704,868
<8x coverage bases	23,534,080	48,176,144	51,596,090
No coverage bases	6,174,731	26,258,294	27,964,065
Annotated <8x coverage bases	1,372	1,348	1,894
HGMD <8x coverage bases	770	777	1,092
ClinVar <8x coverage bases	739	763	1,033
Average coverage	40.45	35.56	35.11
Median coverage	40	35	35
<20x coverage percent	4.82255%	3.95005%	4.38121%
<8x coverage percent	0.80112%	1.63995%	1.75637%
No coverage percent	0.21019%	0.89385%	0.95192%
Annotated <8x percent	0.37112%	0.36463%	0.51232%
HGMD <8x percent	0.37286%	0.37625%	0.52878%
ClinVar <8x percent	0.24882%	0.25690%	0.34781%

The following table provides coverage and sequence depth metrics for three GIAB samples in the coding sequences. This is defined as the sum of all coding sequences in the genome, regardless if targeted or not (as defined by Ensembl).

	NA24149	NA24143	NA12878
Total coverage	35,574,430	35,574,430	35,574,430
Total bases	1,515,714,707	1,334,122,315	1,196,498,356
<20x coverage bases	1,183,332	1,023,561	1,528,704
<8x coverage bases	234,903	344,624	444,046
No coverage bases	36,474	101,797	110,238
Annotated <8x coverage bases	989	972	1,382
HGMD <8x coverage bases	662	673	941
ClinVar <8x coverage bases	460	457	641
Average coverage	42.61	37.50	33.63
Median coverage	43	37	34
<20x coverage percent	3.32636%	2.87724%	4.29720%
<8x coverage percent	0.66031%	0.96874%	1.24822%
No coverage percent	0.10253%	0.28615%	0.30988%
Annotated <8x percent	0.35755%	0.35140%	0.49963%
HGMD <8x percent	0.36345%	0.36949%	0.51662%
ClinVar <8x percent	0.21912%	0.21769%	0.30534%

